

## LOCATION OF GLYCOSYLATION AND ALKYLATION SITES IN ANTHRAQUINONES BY $^1\text{H}$ NMR

SURAJ B. KALIDHAR

Department of Chemistry and Biochemistry, Haryana Agricultural University, Hisar 125 004, India

(Received in revised form 13 January 1989)

**Key Word Index**—Emodin; physcion; morindone; obtusin; dermocybin; anthraquinone; glucoside; gentiobioside; primveroside.

**Abstract**—A comparison of the  $^1\text{H}$  NMR data of the peracetate of emodin with those of emodin-*O*-glycosides helps in the location of the sugar residues. The site of *O*-alkylation in emodin can be similarly determined and the method is useful for other anthraquinones.

### INTRODUCTION

In 1984, we determined the position of the sugar in physcion-8-*O*-gentiobioside [1] with the help of acylation shift data [2]. In this paper, I describe an alternative procedure for locating the position of alkylation or glycosylation in emodin and related compounds, as well as in other anthraquinones.

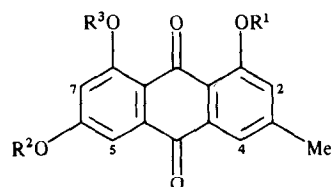
### DISCUSSION

A comparison of the  $^1\text{H}$  NMR data of the peracetates of emodin (1) and emodin-8-*O*-glucoside (2) shows that H-5, the proton which is *para* to the site of glycosylation, undergoes the most significant change in chemical shift (Tables 1 and 2). This change has been measured by the glycosylation shift of the aromatic proton ( $\Delta\text{H}$ ), i.e. the difference between the chemical shifts of the proton in an anthraquinone aglycone peracetate and its glycoside. As is evident from Table 2, H-5 is greatest when the glycoside is at position 8. Similar behaviour has been observed for 8-*O*-alkylation and the shift has been denoted by  $\Delta\text{H-5}$ .

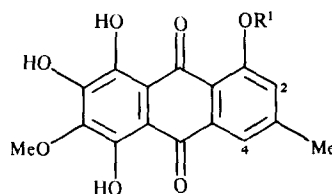
In emodin-1-*O*-glucoside (6), the largest shift is that of H-4, which is *para* to the position of glycosylation (Table 2). In the absence of  $^1\text{H}$  NMR data for physcion-1-*O*-glucoside,\*  $\Delta\text{H-4}$  for dermocybin-1-*O*-glucoside (14), which has a cresol ring, has been included in Table 2 and  $\Delta\text{H-4}$  has been found to be in good agreement with that for emodin-1-*O*-glucoside (6). As expected,  $\Delta\text{H-4}$  is the largest  $\Delta\text{H}$  value when the 1-hydroxyl is alkylated (Tables 1 and 2).

If there is a glycoside at position 6 in emodin (1), it can easily be located in view of the fact that emodin-6-*O*-glycoside does not react with diazomethane. It is also possible to ascertain this position with the help of the proposed method. There is no aromatic proton *para* to the site of glycosylation but it has been observed that the two protons *ortho* to the position of the glycoside, i.e. H-5 and H-7, undergo shifts by more than 0.2 and a similar trend has been observed for 6-*O*-alkylation (Tables 1 and 2).

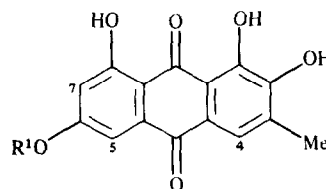
\*There is no mention of  $^1\text{H}$  NMR in the reports [13–15] of physcion-1-*O*-glucoside.



- 1  $\text{R}^1, \text{R}^2, \text{R}^3 = \text{H}$
- 2  $\text{R}^1, \text{R}^2 = \text{H}, \text{R}^3 = \text{glc}$
- 3  $\text{R}^1, \text{R}^2 = \text{H}, \text{R}^3 = \text{gent}$
- 4  $\text{R}^1, \text{R}^2 = \text{H}, \text{R}^3 = \text{prim}$
- 5  $\text{R}^1, \text{R}^2 = \text{H}, \text{R}^3 = \text{glc}$
- 6  $\text{R}^1 = \text{glc}, \text{R}^2, \text{R}^3 = \text{H}$
- 7  $\text{R}^1, \text{R}^2 = \text{H}, \text{R}^3 = \text{Me}$
- 8  $\text{R}^1 = \text{H}, \text{R}^2 = \text{Me}, \text{R}^3 = \text{glc}$
- 9  $\text{R}^1 = \text{H}, \text{R}^2 = \text{Me}, \text{R}^3 = \text{gent}$
- 10  $\text{R}^1 = \text{H}, \text{R}^2 = \text{Me}, \text{R}^3 = \text{prim}$
- 11  $\text{R}^1 = \text{H}, \text{R}^2, \text{R}^3 = \text{Me}$
- 12  $\text{R}^1, \text{R}^2, \text{R}^3 = \text{Me}$
- 13  $\text{R}^1, \text{R}^2 = \text{Me}, \text{R}^3 = \text{H}$



- 14  $\text{R}^1 = \text{H}$
- 15  $\text{R}^1 = \text{glc}$



- 16  $\text{R}^1 = \text{H}$
- 17  $\text{R}^1 = \text{Me}$

Table 1. Chemical shift (in CDCl<sub>3</sub>) of aromatic protons in the <sup>1</sup>H NMR spectra of the peracetates of emodin and related compounds

Peracetate of (br s)	Chemical shift			
	H-2 (br s)	H-4 (br s)	H-5 (d, J = 2.5 Hz)	H-7 (d, J = 2.5 Hz)
Emodin (1) [2]*	7.24	8.03	7.98	7.26
Emodin-8- <i>O</i> -glucoside (2) [2]	7.23	7.97	7.76	7.31
Emodin-8- <i>O</i> -gentiobioside (3) [3]	7.19	7.97	7.78	7.29
Emodin-8- <i>O</i> -primveroside (4) [4]	7.17	7.96	7.78	7.21
Emodin-6- <i>O</i> -glucoside (5) [2]	7.19	7.91	7.54	6.98
Emodin-1- <i>O</i> -glucoside (6) [2]	7.35	7.88	7.97	7.29
6- <i>O</i> -Methylemodin (physcion) and derivatives				
Physcion (7) [2]	7.24	8.03	7.69	6.92
Physcion-8- <i>O</i> -glucoside (8) [2]	7.15	7.90	7.42	6.94
Physcion-8- <i>O</i> -gentiobioside (9) [1]	7.18	7.92	7.50	6.88
Physcion-8- <i>O</i> -primveroside (10) [5]	7.13	7.88	7.45	6.97
8- <i>O</i> -Methylphyscion (11) [2]	7.18	7.96	7.36	6.77
1,8-Di- <i>O</i> -methylphyscion (12) [6]†	7.10	7.64	7.33	6.77
1- <i>O</i> -Methylphyscion (13) [7]	7.05	7.70	7.60	6.95
Anthraquinones having cresol or resorcinol ring				
Dermocybin (14) [2]	7.17	7.86	—	—
Dermocybin-1- <i>O</i> -glucoside (15) [2]	7.24	7.70	—	—
Alaternin (16) [8]	—	8.11	7.95	7.24
6- <i>O</i> -Methylalaternin (17) [9]	—	7.95	7.50	6.80

\*Data from ref. [2] has been reproduced for most of the anthraquinones in this Table.

†No hydroxyl for acetylation in this compound.

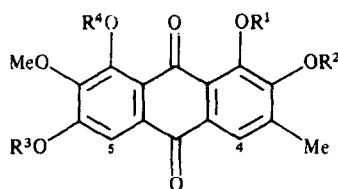
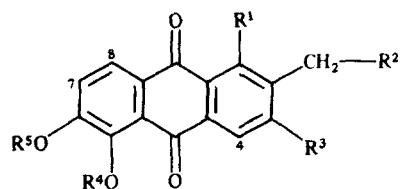
18 R<sup>1</sup>, R<sup>3</sup> = Me, R<sup>2</sup>, R<sup>4</sup> = H19 R<sup>1</sup> = Me, R<sup>2</sup>, R<sup>3</sup>, R<sup>4</sup> = H20 R<sup>1</sup>, R<sup>2</sup> = H, R<sup>3</sup>, R<sup>4</sup> = Me21 R<sup>1</sup>, R<sup>2</sup>, R<sup>4</sup> = H, R<sup>3</sup> = Me22 R<sup>1</sup>, R<sup>2</sup>, R<sup>3</sup>, R<sup>4</sup> = H23 R<sup>1</sup> = OH, R<sup>2</sup>, R<sup>3</sup>, R<sup>4</sup>, R<sup>5</sup> = H24 R<sup>1</sup> = OH, R<sup>2</sup>, R<sup>3</sup>, R<sup>4</sup> = H, R<sup>5</sup> = prim25 R<sup>1</sup> = OH, R<sup>2</sup>, R<sup>3</sup>, R<sup>4</sup> = H, R<sup>5</sup> = rut26 R<sup>1</sup>, R<sup>2</sup>, R<sup>4</sup>, R<sup>5</sup> = H, R<sup>3</sup> = OH27 R<sup>1</sup>, R<sup>2</sup>, R<sup>4</sup> = H, R<sup>3</sup> = OH, R<sup>5</sup> = prim28 R<sup>1</sup>, R<sup>3</sup> = OH, R<sup>2</sup>, R<sup>4</sup>, R<sup>5</sup> = H29 R<sup>1</sup>, R<sup>3</sup> = OH, R<sup>2</sup>, R<sup>4</sup> = H, R<sup>5</sup> = prim30 R<sup>1</sup>, R<sup>2</sup>, R<sup>3</sup> = OH, R<sup>4</sup>, R<sup>5</sup> = H31 R<sup>1</sup>, R<sup>2</sup> = OH, R<sup>3</sup> = -*O*-prim, R<sup>4</sup>, R<sup>5</sup> = H

Table 2. Glycosylation (ΔH)\* or alkylation (Δ'H)† shifts in emodin and related compounds

Peracetates of	ΔH-2 (Δ'H-2)	ΔH-4 (Δ'H-4)	ΔH-5 (Δ'H-5)	ΔH-7 (Δ'H-7)
8- <i>O</i> -Glycosylation (alkylation)				
1, 2	+0.01	+0.06	+0.22	-0.05
1, 3	+0.05	+0.06	+0.20	-0.03
1, 4	+0.07	+0.07	+0.20	+0.05
7, 8	+0.09	+0.13	+0.27	-0.02
7, 9	+0.06	+0.11	+0.19	+0.04
7, 10	+0.11	+0.15	+0.24	-0.05
13, 12	(-0.05)	(+0.06)	(+0.27)	(+0.18)
6- <i>O</i> -Glycosylation (alkylation)				
1, 5	+0.05	+0.12	+0.44	+0.28
1, 7	(0.0)	(0.0)	(+0.29)	(+0.34)
2, 8	(+0.08)	(+0.07)	(+0.34)	(+0.37)
3, 9	(+0.01)	(+0.05)	(+0.28)	(+0.41)
4, 10	(+0.04)	(+0.08)	(+0.33)	(+0.24)
16, 17	—	(+0.16)	(+0.45)	(+0.44)
1- <i>O</i> -Glycosylation (alkylation)				
1, 6	-0.11	+0.15	+0.01	-0.03
14, 15	-0.07	+0.16	—	—
7, 13	(+0.19)	(+0.33)	(+0.09)	(-0.03)
11, 12	(+0.08)	(+0.32)	(+0.03)	(0.0)

\*ΔH = δ value of the aromatic proton in aglycone peracetate minus that in *O*-glycosyl peracetate.†Δ'H = δ value of the aromatic proton in anthraquinone peracetate minus that in *O*-alkyl peracetate.

Table 3. Chemical shift (in CDCl<sub>3</sub>) of aromatic protons in the <sup>1</sup>H NMR spectra of the peracetates of some 2,7-dioxygenated emodins

Peracetate of	Chemical shift	
	H-4 ( <i>br s</i> )	H-5 ( <i>s</i> )
Obtusin (18) [10]	7.95	7.70
Aurantio-obtusin (19) [10]	7.95	7.92
1-Desmethylchryso-obtusin (20) [10]	8.06	7.61
1-Desmethylobtusin (21) [10]	8.03	7.72
1-Desmethyllaurantio-obtusin (22) [10]	8.09	7.94

Table 4. Alkylation shifts of aromatic protons in 2,7-dioxygenated emodins

Peracetates of	Δ' H-4	Δ' H-5
19, 18	0.0	+0.22
21, 20	-0.03	+0.11
22, 21	+0.06	+0.22

Table 5. Chemical shift (in CDCl<sub>3</sub>) of aromatic protons in the <sup>1</sup>H NMR spectra of the peracetates of some 2-methyl (or hydroxymethyl)-anthraquinones

Peracetate of	Chemical shift				
	H-1	H-3	H-4	H-7	H-8
Morindone (23) [11]	—	7.60 <i>d</i>	8.03 <i>d</i>	7.53 <i>d</i>	8.17 <i>d</i> *
Morindone-6- <i>O</i> -primveroside (24) [11]	—	7.62 <i>d</i>	8.03 <i>d</i>	7.38 <i>d</i>	8.19 <i>d</i>
Morindone-6- <i>O</i> -rutinoside (25) [12]	—	7.61 <i>d</i>	8.04 <i>d</i>	7.31†	8.20 <i>d</i>
3,5,6-trihydroxy-2-methyl anthraquinone (26) [11]	8.10 <i>s</i>	—	7.82 <i>s</i>	7.57 <i>d</i>	8.25 <i>d</i>
3,5-dihydroxy-2-methylanthraquinone-6- <i>O</i> -primveroside (27) [11]	8.13 <i>s</i>	—	7.83 <i>s</i>	7.43 <i>d</i>	8.33 <i>d</i>
3-hydroxy morindone (28) [11]	—	—	7.88 <i>s</i>	7.58 <i>d</i>	8.20 <i>d</i>
3-hydroxy morindone-6- <i>O</i> -primveroside (29) [11]	—	—	7.87 <i>s</i>	7.40 <i>d</i>	8.18 <i>d</i>
5,6-dihydroxylucidin (30) [11]	—	—	7.88 <i>s</i>	7.57 <i>d</i>	8.20 <i>d</i>
5,6-dihydroxylucidin-3- <i>O</i> -primveroside (31) [11]	—	—	7.73 <i>s</i>	7.48 <i>d</i>	8.17 <i>d</i>

\**J* = 8–9 Hz.†Overlapped by CHCl<sub>3</sub>.

Consequently, for a glycoside, if ΔH-4 gives the highest value then 1-*O*-glycosylation is indicated and when ΔH-5 and ΔH-7 are both more than 0.2, 6-*O*-glycosylation is indicated. When 6-*O*-glycosylation is absent, a large value for ΔH-5 suggests 8-*O*-glycosylation. The position of *O*-alkylation can be similarly determined in emodin and physcion.

A comparison of the peracetates of aurantio-obtusin (19) and obtusin (18) shows a higher value for ΔH-5 than for ΔH-4 (Tables 3 and 4). In the absence of a proton *para* to position 6 and in the presence of only one *ortho* proton (H-5), a higher value of ΔH-5 is in accordance with the expected trend. The other alkylation shifts shown in Tables 3 and 4 are consistent with the proposed method.

A comparison of the peracetates of morindone (23) and morindone-6-*O*-primveroside (24) (Table 5) shows the greatest shift for H-7, in accordance with the expected trend. The other glycosylation shifts shown in Table 6 are also consistent with the proposed method.

## REFERENCES

- Kalidhar, S. B. and Sharma, P. (1984) *Phytochemistry* **23**, 1196.
- Steglich, W. and Lösel, W. (1969) *Tetrahedron* **25**, 4391.
- Demuth, G., Hinzo, H., Selligmann, O. and Wagner, H. (1978) *Planta Med.* **33**, 53.
- Rauwald, H. W. (1983) *Z. Naturforsch.* **38C**, 170.
- Coskun, M., Sakushima, A., Kitagawa, S. and Nishibe, S. (1984) *Phytochemistry* **23**, 1485.
- Thomson, R. H. (1971) *Naturally Occurring Quinones* 2nd Edn., p. 74. Academic Press, London.
- Murakami, T., Ikeda, K. and Takido, M. (1968) *Chem. Pharm. Bull.* **16**, 2299.
- Kitanaka, S., Kimura, F. and Takido, M. (1985) *Chem. Pharm. Bull.* **33**, 1274.

Table 6. Glycosylation shifts of aromatic protons in 2-methyl (or hydroxymethyl)-anthraquinones

Peracetates of	ΔH-1	ΔH-3	ΔH-4	ΔH-7	ΔH-8
23, 24	—	-0.02	0.0	+0.15	-0.02
23, 25	—	-0.01	-0.01	+0.22	-0.03
26, 27	-0.03	—	-0.01	+0.14	-0.08
28, 29	—	—	+0.01	+0.18	+0.02
30, 31	—	—	+0.15	+0.09	+0.03

9. Kalidhar, S. B. and Sharma, P. (1985) *J. Indian Chem. Soc.* **LXII**, 411.
10. Kitanaka, S. and Takido, M. (1984) *Chem. Pharm. Bull.* **32**, 860.
11. Inoue, K., Nayeshiro, H., Inouye, H. and Zenk, M. (1981) *Phytochemistry* **20**, 1693.
12. Vermes, B., Farkas, L. and Wagner, M. (1980) *Phytochemistry* **19**, 119.
13. Lal, J. and Gupta, P. C. (1973) *Experientia* **29**, 141.
14. Niranjana, G. S. and Gupta, P. C. (1973) *Planta Med.* **23**, 298.
15. Smith, R. M. and Ali, S. (1979) *N. Z. J. Sci.* **22**, 123.